



Full wwPDB NMR Structure Validation Report ⓘ

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PDB ID : 1XR0
Title : Structural Basis of SNT PTB Domain Interactions with Distinct Neurotrophic Receptors
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This is a Full wwPDB NMR Structure Validation Report for a publicly released PDB entry.

We welcome your comments at validation@mail.wwpdb.org

A user guide is available at

<https://www.wwpdb.org/validation/2017/NMRValidationReportHelp>

with specific help available everywhere you see the ⓘ symbol.

The types of validation reports are described at <http://www.wwpdb.org/validation/2017/FAQs#types>.

The following versions of software and data (see [references ⓘ](#)) were used in the production of this report:

MolProbity : 4.02b-467
Percentile statistics : 20191225.v01 (using entries in the PDB archive December 25th 2019)
RCI : v_1n_11_5_13_A (Berjanski et al., 2005)
PANAV : Wang et al. (2010)
ShiftChecker : 2.29
Ideal geometry (proteins) : Engh & Huber (2001)
Ideal geometry (DNA, RNA) : Parkinson et al. (1996)
Validation Pipeline (wwPDB-VP) : 2.29

1 Overall quality at a glance

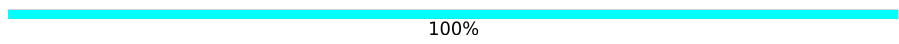
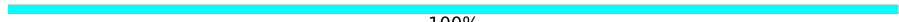
The following experimental techniques were used to determine the structure:

SOLUTION NMR

The overall completeness of chemical shifts assignment was not calculated.

There are no overall percentile quality scores available for this entry.

The table below summarises the geometric issues observed across the polymeric chains and their fit to the experimental data. The red, orange, yellow and green segments indicate the fraction of residues that contain outliers for ≥ 3 , 2, 1 and 0 types of geometric quality criteria. A cyan segment indicates the fraction of residues that are not part of the well-defined cores, and a grey segment represents the fraction of residues that are not modelled. The numeric value for each fraction is indicated below the corresponding segment, with a dot representing fractions $\leq 5\%$

Mol	Chain	Length	Quality of chain
1	A	22	 100%
2	B	129	 100%

2 Ensemble composition and analysis

This entry contains 1 models. Identification of well-defined residues and clustering analysis are not possible.

3 Entry composition

There are 2 unique types of molecules in this entry. The entry contains 2436 atoms, of which 1211 are hydrogens and 0 are deuteriums.

- Molecule 1 is a protein called Basic fibroblast growth factor receptor 1.

Mol	Chain	Residues	Atoms						Trace
			Total	C	H	N	O	S	
1	A	22	368	108	194	36	29	1	0

- Molecule 2 is a protein called FGFR signalling adaptor SNT-1.

Mol	Chain	Residues	Atoms						Trace
			Total	C	H	N	O	S	
2	B	129	2068	649	1017	190	205	7	0

There are 3 discrepancies between the modelled and reference sequences:

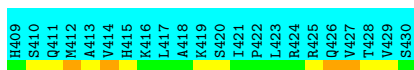
Chain	Residue	Modelled	Actual	Comment	Reference
B	8	MET	-	cloning artifact	UNP Q8WU20
B	9	GLY	-	cloning artifact	UNP Q8WU20
B	10	SER	-	cloning artifact	UNP Q8WU20

4 Residue-property plots [i](#)

These plots are provided for all protein, RNA, DNA and oligosaccharide chains in the entry. The first graphic is the same as shown in the summary in section 1 of this report. The second graphic shows the sequence where residues are colour-coded according to the number of geometric quality criteria for which they contain at least one outlier: green = 0, yellow = 1, orange = 2 and red = 3 or more. Stretches of 2 or more consecutive residues without any outliers are shown as green connectors. Residues which are classified as ill-defined in the NMR ensemble, are shown in cyan with an underline colour-coded according to the previous scheme. Residues which were present in the experimental sample, but not modelled in the final structure are shown in grey.

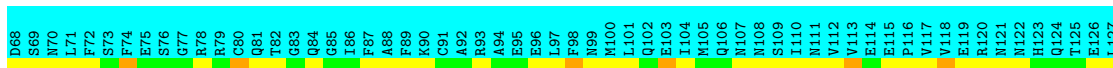
- Molecule 1: Basic fibroblast growth factor receptor 1

Chain A:  100%



- Molecule 2: FGFR signalling adaptor SNT-1

Chain B:  100%



5 Refinement protocol and experimental data overview

The models were refined using the following method: *Structures of the SNT-1 PTB domain in complex with the hFGFR1 peptide were calculated with a distance geometry and simulated annealing protocol by using the X-PLOR program ([4]). NOE distance and dihedral angle restraints were treated with a square-well potential of 50 kcal mol⁻¹. A total of 2448 manually assigned NOE-derived distance restraints were obtained from the ¹⁵N- or ¹³C-edited NOESY data. Included in this figure are 251 intrapeptide and 258 intermolecular distance restraints. Additionally, 255 unambiguous and 52 ambiguous distance restraints were identified from the NOE data by using ARIA. The final structure calculations employed a total of 2755 NOE restraints obtained from the manual and the ARIA-assisted assignments, 2703 of which were unambiguously assigned NOE-derived distance restraints that comprise 1072 intraresidue, 466 sequential, 216 medium-range, and 949 long-range NOEs. In addition, 70 hydrogen-bond distance restraints for 35 hydrogen bonds and 19 ϕ -angle restraints were also used in the structure calculations. For the ensemble of the final 20 structures, no distance or torsional angle restraint was violated by more than 0.4 or 5, respectively. The distance-violation, dihedral-violation, and total energies were 74.4 1.7 kcal mol⁻¹, 0.82 0.08 kcal mol⁻¹, and 262.0 6.0 kcal mol⁻¹, respectively. The Lennard-Jones potential, which was not used during any refinement stage, was 659.3 23.1 kcal mol⁻¹ for the final structures. Ramachandran plot analysis by Procheck-NMR showed that in the final structures of the complex, 98.1% of the backbone geometries of the non-Gly and non-Pro residues in the complex (protein residues 18-116 and peptide residues 412-430) and nearly 100% in the secondary structure (protein residues 19-24, 35-40, 45-49, 52-57, 63-68, 71-76, 85-90, 94-107, and 111-115 and peptide residues 426-430) lie within energetically favorable or allowed regions..*

Of the 100 calculated structures, 1 were deposited, based on the following criterion: *structures with the least restraint violations.*

The following table shows the software used for structure solution, optimisation and refinement.

Software name	Classification	Version
X-PLOR	structure solution	3.851
ARIA	refinement	1.1

No chemical shift data was provided.

6 Model quality [i](#)

6.1 Standard geometry [i](#)

There are no covalent bond-length or bond-angle outliers.

There are no bond-length outliers.

There are no bond-angle outliers.

There are no chirality outliers.

There are no planarity outliers.

6.2 Too-close contacts [i](#)

In the following table, the Non-H and H(model) columns list the number of non-hydrogen atoms and hydrogen atoms in each chain respectively. The H(added) column lists the number of hydrogen atoms added and optimized by MolProbity. The Clashes column lists the number of clashes averaged over the ensemble.

Mol	Chain	Non-H	H(model)	H(added)	Clashes
1	A	0	0	0	0
2	B	0	0	0	0
All	All	0	0	0	-

The all-atom clashscore is defined as the number of clashes found per 1000 atoms (including hydrogen atoms). The all-atom clashscore for this structure is -.

There are no clashes.

6.3 Torsion angles [i](#)

6.3.1 Protein backbone [i](#)

In the following table, the Percentiles column shows the percent Ramachandran outliers of the chain as a percentile score with respect to all PDB entries followed by that with respect to all NMR entries. The Analysed column shows the number of residues for which the backbone conformation was analysed and the total number of residues.

Mol	Chain	Analysed	Favoured	Allowed	Outliers	Percentiles
1	A	0	-	-	-	-
2	B	0	-	-	-	-
All	All	0	-	-	-	-

There are no Ramachandran outliers.

6.3.2 Protein sidechains [i](#)

In the following table, the Percentiles column shows the percent sidechain outliers of the chain as a percentile score with respect to all PDB entries followed by that with respect to all NMR entries. The Analysed column shows the number of residues for which the sidechain conformation was analysed and the total number of residues.

Mol	Chain	Analysed	Rotameric	Outliers	Percentiles
1	A	0	-	-	-
2	B	0	-	-	-
All	All	0	-	-	-

There are no protein residues with a non-rotameric sidechain to report.

6.3.3 RNA [i](#)

There are no RNA molecules in this entry.

6.4 Non-standard residues in protein, DNA, RNA chains [i](#)

There are no non-standard protein/DNA/RNA residues in this entry.

6.5 Carbohydrates [i](#)

There are no monosaccharides in this entry.

6.6 Ligand geometry [i](#)

There are no ligands in this entry.

6.7 Other polymers [i](#)

There are no such molecules in this entry.

6.8 Polymer linkage issues [i](#)

There are no chain breaks in this entry.

7 Chemical shift validation

No chemical shift data were provided